MAP KINASE PHOSPHATASE1 and PROTEIN TYROSINE PHOSPHATASE1 Are Repressors of Salicylic Acid Synthesis and SNC1-Mediated Responses in Arabidopsis

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Mitogen-activated protein (MAP) kinase phosphatases are important negative regulators of the levels and kinetics of MAP kinase activation that modulate cellular responses. The dual-specificity phosphatase MAP KINASE PHOSPHATASE1 (MKP1) was previously shown to regulate MAP KINASE6 (MPK6) activation levels and abiotic stress responses in Arabidopsis thaliana. Here, we report that the mkp1 null mutation in the Columbia (Col) accession results in growth defects and constitutive biotic defense responses, including elevated levels of salicylic acid, camalexin, PR gene expression, and resistance to the bacterial pathogen Pseudomonas syringae. PROTEIN TYROSINE PHOSPHATASE1 (PTP1) also interacts with MPK6, but the ptp1 null mutant shows no aberrant growth phenotype. However, the pronounced constitutive defense response of the mkp1 ptp1 double mutant reveals that MKP1 and PTP1 repress defense responses in a coordinated fashion. Moreover, mutations in MPK3 and MPK6 distinctly suppress mkp1 and ptp1 phenotypes, indicating that MKP1 and PTP1 act as repressors of inappropriate MPK3/MPK6-dependent stress signaling. Finally, we provide evidence that the natural modifier of mkp1 in Col is largely the disease resistance gene homolog SUPPRESSOR OF npr1-1, CONSTITUTIVE 1 (SNC1) that is absent in the Wassilewskija accession. Our data thus indicate a major role of MKP1 and PTP1 in repressing salicylic acid biosynthesis in the autoimmune-like response caused by SNC1.

INTRODUCTION

Mitogen-activated protein (MAP) kinase cascades are conserved signal transduction systems in eukaryotic cells that transmit and integrate a multitude of environmental and intracellular signals. These cascades relay signals by sequential phosphorylation and consist of three interlinked kinase components, a MAP kinase kinase kinase (MAPKKK), a MAP kinase kinase (MAPKK), and a terminal MAP kinase (MAPK). MAPKs are activated by dual phosphorylation of Thr and Tyr within their T-X-Y consensus sequence by the dual-specificity MAPKKs. The magnitude and duration of MAPK activation determines the outcome of the cellular reaction (Marshall, 1995; Keyse, 2008). Therefore, tight regulation of MAPK activation is a prerequisite for producing specific and adequate physiological responses (McClellan et al., 2007). Deregulation of MAPK activity has a broad range of deleterious effects in all eukaryotic organisms, including being a common alteration in human cancers (Wu, 2007).

Regulated dephosphorylation and inactivation of MAPKs by protein phosphatases directly counterbalances activation through MAPKKs and is a major determinant of the physiological outcome of signaling (e.g., Junttila et al., 2008; Keyse, 2008). Since phosphorylation of both Thr and Tyr residues is required for MAPK activity, dephosphorylation of either amino acid is sufficient for inactivation. This can be achieved by members of the three major classes of protein phosphatases, namely, Tyr-specific phosphatases (PTPs), Ser/Thr-specific phosphatases, or dual-specificity (Thr/Tyr) protein phosphatases (DSPs) (Keyse, 2008). In particular, members of a subgroup of the DSP class are known to be dedicated MAPK phosphatases (MKPs) (Camps et al., 2000). Defects in such dual-specificity MKPs have a broad range of detrimental effects in multicellular eukaryotes (e.g., Martin-Blanco et al., 1998; Ulm et al., 2001; Monroe-Augustus et al., 2003; Naoi and Hashimoto, 2004; Zhang et al., 2004; Christie et al., 2005; Lee and Ellis, 2007).
In *Arabidopsis thaliana*, members of all three major classes of protein phosphatases were linked to MAPK deactivation, albeit to different degrees. A PP2C-type Ser/Thr phosphatase, ABI1, was found to interact with and inactivate MAP KINASE6 (MPK6), implicating it in abscisic acid–dependent stress signaling (Leung et al., 2006). Another PP2C-type phosphatase, AP2C1, was recently shown to inactivate wound stress–responsive MPK4 and MPK6 in vivo, and its overexpression was found to compromise innate immunity against a necrotrophic fungal pathogen (Schweighofer et al., 2007). Mutant ap2c1 plants, on the other hand, were not significantly affected in their pathogen response; however, they were shown to have an elevated wound response and to be more resistant to herbivorous attack by spider mites (Schweighofer et al., 2007).

PTP1 is the only member of the phosphotyrosine-specific protein phosphatases in *Arabidopsis*. *PTP1* gene expression was found to be altered in response to different environmental stresses (Xu et al., 1998). PTP1 was shown to be able to dephosphorylate and deactivate MPK4 and MPK6 in vitro, and an involvement of PTP1 in oxidative stress signaling was suggested (Huang et al., 2000; Gupta and Luan, 2003), but its role in vivo has not yet been reported.

Five DSP-type phosphatases with described or predicted MKP function are encoded in the genome of *Arabidopsis*. For one of them, the dual-specificity protein Tyr phosphatase 1 (DsPTP1; Gupta et al., 1998), no in vivo role has been described yet. The other four DSP-type MKPs were genetically linked to abscisic acid/auxin signaling (INDOLE-3-BUTYRIC ACID RESPONSE5; Monroe-Augustus et al., 2003; Lee et al., 2009), microtubule organization (PROPYZAMIDE-HYPERSENSITIVE1; Naoi and Hashimoto, 2004), oxidative stress tolerance (MKP2; Lee and Ellis, 2007), and genotoxic and salt stress tolerance (MKP1; Ulm et al., 2001, 2002).

MKP1 is required for genotoxic stress resistance in *Arabidopsis* and was found to interact with the stress-activated MPK3, MPK4, and MPK6 in a directed yeast two-hybrid assay (Ulm et al., 2001, 2002). MKP1 was further shown to regulate the genotoxic stress-responsive activity level of MPK6 in vivo (Ulm et al., 2002). The *mkp1* mutant, however, does not show general stress sensitivity, as, on the contrary, *mkp1* was found to be more tolerant to salt stress (Ulm et al., 2002). Recently, it was described that MKP1 is regulated by calmodulin (CaM) through two CaM binding domains. Binding of CaM increased the phosphatase activity of *Arabidopsis* MKP1 about twofold, indicating a link between Ca$^{2+}$ and MKP1-mediated regulation of MAPK signaling (Lee et al., 2008). The tobacco (*Nicotiana tabacum*) MKP1, however, was not activated by CaM binding but by interaction with its substrate, the SALICYLIC ACID-INDUCED PROTEIN KINASE (SIPK), which is the ortholog of *Arabidopsis* MPK6 (Yamakawa et al., 2004; Katou et al., 2005). Transient overexpression of *N. tabacum* MKP1 in tobacco leaves abolished SIPK-mediated cell death (Katou et al., 2005). The rice (*Oryza sativa*) MKP1 was recently described as a negative regulator of wound responses, and the rice insertional mutant line *mkp1* shows a semidwarf phenotype (Katou et al., 2007).

Despite the importance of MAPK pathways in a broad range of stress responses and developmental programs, the contribution of negative regulatory MKPs has been described genetically only in very few cases in multicellular eukaryotes. Similarly, the specific and/or overlapping functions of different classes of MAPK-regulating phosphatases are not well understood. Here, we show that *Arabidopsis* MKP1 and PTP1 have redundant functions in the suppression of salicylic acid (SA) and camalexin biosynthesis and defense responses. We found that the constitutive defense response in *mkp1* and the *mkp1 ptp1* double mutant is partially dependent on MPK3 and MPK6 and accumulation of SA. Moreover, the described phenotype of *mkp1* and *mkp1 ptp1* is only apparent in the Columbia (Col) ecotype, but not in the Wassilewskija (Ws) ecotype. Previous work has identified the Toll Interleukin 1 receptor/nucleotide binding/leucine-rich repeat (TIR-NB-LRR) receptor-like resistance gene homolog *SUPPRESSOR OF npr1-1, CONSTITUTIVE1 (SNC1)*, which induces constitutive defense responses and dwarfs growth when overexpressed or hyperactivated by mutation (Stokes et al., 2002; Zhang et al., 2003; Yang and Hua, 2004; Li et al., 2007). This gene is specific to Col, and there is no closely related ortholog in Ws (Yang and Hua, 2004). Our genetic analysis indeed identified *SNC1* as a natural modifier of MKP1 function.

**RESULTS**

**MKP1 and PTP1 Interact with and Dephosphorylate MPK6**

PTP1 was previously shown to dephosphorylate activated MPK6 in vitro (Gupta and Luan, 2003). However, we could not identify direct interaction of PTP1 with MPK6 or other tested MKPs in a directed yeast two-hybrid assay. A possible reason for this might be a rather transient and weak interaction of PTP1 with its substrate. To investigate direct interaction in planta, we used the bimolecular fluorescence complementation (BiFC) assay, which stabilizes a protein–protein interaction after the reconstitution of the C-terminal and N-terminal parts of a split yellow fluorescent protein (YFP) (Kerppola, 2006). Indeed, YN-PTP1 was found to interact with itself (YC-PTP1), YC-MPK6, and to a lesser extent YC-MPK3 in transiently transformed mustard (*Sinapis alba*) hypocotyl cells (Figure 1A), a well-established system for BiFC assays in planta (e.g., Stolpe et al., 2005; Favory et al., 2009). The YFP fluorescence resulting from cobombardment of YN-PTP1 and YC-MPK4 is very weak; thus, their direct physical interaction remains uncertain (Figure 1A). No sign of interaction could be detected between YN-PTP1 and YC-MPK1, YC-MPK2, or any of the empty vector controls (Figure 1A). Moreover, we could confirm the yeast two-hybrid based interaction of MKP1 with MPKs 3, 4, and 6 (Ulm et al., 2002) in the BiFC assays (Figure 1A). Using a transient protoplast expression system, we could show that MKP1 is able to inactivate coexpressed and activated MPK6 (Figure 1B). However, we could not express PTP1 to detectable levels in the protoplast system to convincingly assay its effect on MPK6 activity.

Interestingly, the interaction of YC-MPK6 with YN-MKP1 was found to be primarily cytoplasmic, whereas the interaction of YC-MPK6 with YN-PTP1 was primarily nuclear (Figure 1A). We thus generated stable transgenic lines expressing MKP1-YFP and PTP1-YFP driven by the cauliflower mosaic virus 35S promoter in *mkp1* and *ptp1* mutant backgrounds, respectively. In agreement
with the BiFC interaction data, confocal microscopy suggests that MKP1-YFP is mainly localized to the cytosol, whereas PTP1-YFP can be detected both in the cytosol and nucleus (Figure 1C).

**MKP1 and PTP1 Perform Partially Redundant Functions in Vivo**

The overlap of their interaction partners indicated functional redundancy of MKP1 and PTP1. To directly test this possibility, we isolated a T-DNA insertion line disrupting the **PTP1** gene (**ptp1-1**, SALK_118658). In this line, the T-DNA is inserted into exon 4 and results in the absence of wild-type **PTP1** mRNA, indicating that it is a functional null mutant (see Supplemental Figure 1 online). The **ptp1** mutant, however, did not show any obvious change in its morphology or development under standard growth conditions (Figure 2A; see Supplemental Figure 1 online).

As the only available **mkp1** null mutant was in the Ws background (Ulm et al., 2001), we introgressed this allele into the Col accession before crossing it with the **ptp1** mutant. Surprisingly, the introgression of **mkp1** into Col itself generated plants with altered morphology and development not seen in the **mkp1** (Ws) mutant. The **mkp1**(Col) plants showed weak dwarfism, aberrant leaf development, long pedicels, early senescence, stunted and misshaped inflorescences, and low fertility (Figure 2A; see Supplemental Figure 2A online). This mutant phenotype was (1) apparent even after 15 backcrosses to Col, (2) complemented by expression of Polyoma epitope- and YFP-tagged MKP1 (see Supplemental Figures 2B and 2C online; data not shown), and (3) supported by a comparable phenotype of a cosuppression line.
isolated among ProMKP1::GUS transgenic lines (see Supplemental Figures 2D to 2F online). From now on in the text, we will differentiate between mkp1(Col) and mkp1(Ws) for the single mutants; all double mutants were generated using mkp1(Col).

We then crossed the ptpl and mkp1(Col) mutants. Strikingly, the mkp1 ptp1 double mutant plants exhibit a severe dwarf phenotype (Figure 2A). These plants show numerous dramatic developmental defects, including early senescence and infertility. The mkp1(Col) and mkp1 ptp1 growth phenotypes become apparent about 2 weeks after germination and are not established at the earlier seedling stage (see Supplemental Figure 3 online). Altogether, we conclude that MKP1 and PTP1 have partially redundant functions under standard growth conditions, with MKP1 having the predominant role.

MPK3 and MPK6 Mediate mkp1 and mkp1 ptp1 Developmental Defects

As both MKP1 and PTP1 regulate MPK6 and likely MPK3, deregulation of MPK6 and/or MPK3 might be the underlying cause for the aberrant phenotype of mkp1(Col) and mkp1 ptp1. Consistently, the double mkp1 mpk6-2 and triple mkp1 ptp1 mpk6-2 knockout mutants show a significantly improved growth phenotype compared with mkp1(Col) and mkp1 ptp1, respectively, although overall plant size was still slightly reduced and early senescence remained (Figure 2A). It should be noted that suppression was very similar with an independent mpk6 allele (mpk6-3; see Supplemental Figure 4 online).

Similarly to mpk6, the mpk3 null mutation partially suppressed the mkp1(Col) phenotype in mkp1 mpk3 double mutants (Figure 2A). Interestingly, however, MPK3 and MPK6 contributed differently to the mkp1(Col) phenotype: (1) mpk3 suppressed the early senescence phenotype of mkp1, whereas mpk6 did not; (2) mpk6 suppressed the inflorescence morphology phenotype of mkp1, whereas mpk3 did not; and (3) mpk6 suppressed the decreased fertility phenotype of mkp1, whereas mpk3 did not. Thus, mutations in mpk3 and mpk6 suppress mkp1 phenotypes, although distinctly and to a different extent.

Protein gel blot analysis using specific antibodies confirmed the absence of MPK3 and MPK6 in the respective lines and indicated that MPK3 and MPK4 protein levels are slightly elevated in mkp1(Col) and mkp1 ptp1 compared with wild-type plants (Figure 2B). More importantly, immunoblot analysis using an antiphospho-p44/42 antibody, which recognizes only the
doubly phosphorylated and thus active forms of MPK3 and MPK6, identified elevated levels of the phosphorylated forms of these two kinases in mkp1(Col) and mkp1 ptp1 compared with the wild type under standard growth conditions (Figure 2C). As expected, in mkp1 mpk3 and mkp1 mpk6, the cross-reacting bands of phospho-MPK6 and phospho-MPK3 are absent, respectively (Figure 2C). Thus, the loss-of MKP1 and MKP1/PTP1 is associated with increased levels of active MPK3 and MPK6, and in agreement, the mkp1(Col) and mkp1 ptp1 phenotypes largely depend on the presence of MPK3 and MPK6. We thus conclude that loss of MKP1 and combined loss of MKP1 and PTP1 lead to a multitude of developmental and morphological alterations, several of which could be genetically attributed to MPK3 and/or MPK6 function.

**PR Gene Expression Is Elevated in mkp1 and mkp1 ptp1**

The phenotypes of mkp1(Col) and mkp1 ptp1 indicate a constitutive stress response, also displayed by mutants with a constitutive pathogen response (e.g., Jirage et al., 2001). This phenotype is often associated with an upregulation of PATHOGENESIS-RELATED (PR) gene expression. Quantitative real-time PCR analysis of PR1 and PR5 transcript abundance in 22-d-old soil-grown plants revealed an up to 25-fold increase of PR1 and 6-fold increase of PR5 transcript levels in mkp1 ptp1 compared with wild-type Col (Figures 2D and 2E). The absence of MKP6 partially suppresses this molecular phenotype of mkp1(Col) and mkp1 ptp1 as well (Figures 2D and 2E). This result is in agreement with partial suppression of growth and developmental aberrations of mkp1 ptp1. Similarly, mpk3 knockout in the mkp1(Col) background reduced PR1 and PR5 transcription back to wild-type levels (Figures 2D and 2E). We conclude that the growth phenotype and PR gene hyperexpression indicate an MPK3/MPK6-dependent, constitutively elevated stress response in mkp1 ptp1 and to a lesser extent in mkp1(Col).

**SA Accumulates to High Levels in mkp1 and mkp1 ptp1**

A high level of PR gene expression is often associated with elevated levels of the phytohormone SA. Therefore, we determined the SA levels in rosette leaves of 22-d-old plants grown under standard conditions in soil. In agreement with our phenotypic and molecular analysis, mkp1(Col) shows a 5-fold accumulation of total SA compared with the wild type, and the mkp1 ptp1 double mutant shows a 13-fold increase (Figure 3A). In both cases, the SA hyperaccumulation was dependent on functional mpk6; thus, SA levels were comparable to those in the wild type in mkp1 mpk6 and mkp1 ptp1 mpk6 mutant lines. Similarly, the mkp1 mpk3 double mutant resulted in SA levels comparable to those in the wild type (Figure 3A). It should be noted that separate analysis of free and conjugated SA levels in both cases agreed with the data of total SA (see Supplemental Figure 5 online). To identify a possible molecular cause for the differences in SA levels, we determined the expression levels of the ICS1/SID2 gene, which encodes the isochorismate synthase of the major SA biosynthetic pathway (Wildermuth et al., 2001). In agreement with the phenotypes, PR gene expression as well as SA measurements, we identified a corresponding overexpression of ICS1 in mkp1(Col) and mkp1 ptp1 that was dependent on MPK3 and MPK6 (Figure 3B).

**Elevated SA Levels Are Responsible for the mkp1 and mkp1 ptp1 Phenotypes**

To determine the contribution of SA hyperaccumulation to the mkp1(Col) and mkp1 ptp1 phenotypes, we crossed these mutants with a NahG transgenic line expressing a bacterial salicylate hydroxylase that degrades SA in plants (Gaffney et al., 1993). Indeed, expression of NahG dramatically reduced SA levels and largely suppressed the mkp1(Col) and mkp1 ptp1 phenotypes, including PR gene expression (Figures 3A, 3C, and 3D; see Supplemental Figures 5A and 5B online). At later developmental stages, however, a number of phenotypes remained in the NahG suppressor lines, including early leaf senescence, a weak florescence phenotype, and infertility. Interestingly, NahG expression also reduced MPK3, MPK4, and MPK6 protein levels, indicating that SA modulates their accumulation (Figure 3B).

The lipase-like proteins ENHANCED DISEASE SUSCEPTIBILITY1 (EDS1) and PHYTOALEXIN DEFICIENT4 are important regulators of SA synthesis in plant defense signaling (Wiermer et al., 2005). Therefore, we tested if mutations in these two factors are also able to suppress the mkp1(Col) phenotype, which was indeed found to be the case (Figure 3E).

We thus conclude that the phenotypes of mkp1(Col) and mkp1 ptp1 are largely due to increased SA synthesis and accumulation, indicating that a major function of MKP1 and PTP1 is to repress defense responses potentially associated with increased bacterial growth (Fan et al., 2008). Indeed, in comparison to the wild type, mkp1(Col) allowed much less growth of *P. syringae* pv *tomato DC3000* strain carrying a chromosomal insertion of the luxCDABE operon from *Photorhabdus luminescens*, encoding both a luciferase reporter (*luxAB*) and enzymes for the production of its substrate (*luxCDE*), under a constitutive promoter to noninvasively determine bacterial growth (Fan et al., 2008). Indeed, in comparison to the wild type, mkp1(Col) allowed much less growth of *P. syringae* as determined by bioluminescence, which reflects bacterial growth in the infected *Arabidopsis* leaves (Figure 4A), and leaf bacteria measurement by serial dilution plating (Figure 4B). This enhanced pathogen resistance of mkp1(Col) required MPK6, as mkp1 mpk6 were clearly less resistant than mkp1(Col) and more similar to the wild type (Figure 4C). Surprisingly, however, pathogen resistance of mkp1 mpk3 was similar to that of mkp1 (Figure 4C). Thus, it seems that the elevated pathogen resistance of mkp1 requires MPK6 but not MPK3 and that resistance is not strictly correlated with elevated SA levels and PR gene expression.

Camalexin is an important phytoalexin in *Arabidopsis* often associated with pathogen resistance. Moreover, it was previously shown that constitutive activation of MPK3 and MPK6
Figure 3. SA Accumulation Is Largely Responsible for the Aberrant \textit{mkp1}(Col) and \textit{mkp1 ptp1} Phenotypes.

(A) Total SA levels of 22-d-old soil-grown plants, as determined by HPLC. The average of three independent biological samples is shown for each genotype. Error bars represent SD.

(B) Quantitative RT-PCR analysis of \textit{ICS1} expression levels. Representative data of three biological replicates are shown. Error bars represent SD of the technical triplicates.

(C) Photographs of 40-d-old soil-grown plants showing that expression of NahG largely suppresses the \textit{mkp1 ptp1} growth phenotype (note that the Col and \textit{mkp1 ptp1} photographs are identical to those shown in Figure 2A; all photographs were taken in parallel, under identical conditions). Bars = 2 cm.

(D) Quantitative RT-PCR analysis of \textit{PR1} and \textit{PR5} expression levels in 22-d-old \textit{mkp1 ptp1} and \textit{mkp1 ptp1 NahG} compared with wild-type Col. Representative data of three biological replicates are shown. Error bars represent the SD of the technical triplicates.

(E) Photographs of 22-d-old plants grown on soil under standard conditions showing that \textit{eds1} and \textit{pad4} mutations suppress the \textit{mkp1} growth phenotype. Bars = 2 cm.

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elicits accumulation of camalexin and thereby strengthens pathogen resistance (Ren et al., 2008). Thus, we determined camalexin levels in mkp1(Col) and mkp1 ptp1 under standard growth conditions. In agreement with our previous results, we found ~40-fold increases in camalexin levels in mkp1(Col) and 600-fold in mkp1 ptp1 that are dependent on the presence of MPKs 3 and 6 (Figure 4D). Moreover, the absence of camalexin in the mkp1 ptp1 NahG line is consistent with the requirement of SA for its biosynthesis and the reported effect of NahG on camalexin accumulation (Zhao and Last, 1996; Heck et al., 2003) (Figure 4D).

**The Col Accession-Specific Receptor-Like R Gene Homolog, SNC1, Is a Natural Modifier of mkp1**

The phenotypes that we describe for mkp1(Col) were reminiscent of the previously described bonsai 1 (bon1) mutation in a copine family member (Hua et al., 2001). In particular, similar to bon1 (Yang and Hua, 2004), the mkp1(Col) growth phenotype is (1) restricted to the Col accession and not apparent in Ws, (2) associated with elevated levels of SA and PR gene expression, (3) suppressed by mutant combination with eds1, pad4, and...
NahG, and (4) suppressed by elevated temperature (see Supplemental Figure 6 online). Importantly, Yang and Hua (2004) could show that the bon1-1 loss-of-function mutation activates the Col accession-specific TIR-NB-LRR receptor-like SNC1, leading to constitutive defense responses and, consequently, reduced cell growth. Our data indicate a related role of MKP1 in regulating plant growth homeostasis by repressing inappropriate stress signaling. We thus hypothesized that the Col-specific SNC1 resistance gene homolog is responsible for the constitutive defense response apparent in mkp1(Col). To test this, we generated an mkp1 snc1-11 double mutant; indeed, the snc1 mutation could partially suppress the mkp1(Col) phenotype: both growth habit and PR gene expression are more similar to those of the wild type than to those of the single mkp1(Col) mutant (Figures 5A and 5B). It is also of note that the snc1 single mutant exhibited PR1 and PR5 expression levels lower than those in the wild type, indicating a low-level SNC1-mediated defense response in wild-type Col plants (Figure 5B).

Previously, it was shown that the SNC1-dependent constitutive defense responses and dwarf phenotype are associated with transcriptional upregulation of SNC1 in bon1 mutants as well as in the epigenetic variant bal (Stokes et al., 2002; Yang and Hua, 2004; Li et al., 2007). Similarly, mutational hyperactivation and transgenic overexpression of SNC1 leads to an SA-dependent constitutive defense response and dwarfed growth (Stokes et al., 2002; Zhang et al., 2003; Li et al., 2007). Therefore, we tested SNC1 expression levels in mkp1(Col) but could not identify a statistically significant upregulation (Figure 5C). However, under our growth conditions, we could also not detect a significant increase of SNC1 expression in bon1-1 (Figure 5C), even though it showed a strong growth phenotype. Similarly, BON1 mRNA levels were not strongly affected in mkp1(Col) (Figure 5D).

The overlapping functions of MKP1 and PTP1 described in this work indicated that PTP1 might also be involved in SNC1-mediated responses. To test this, we generated an mkp1 ptp1 snc1-11 triple mutant. As expected, the snc1 mutation could partially suppress the mkp1 ptp1 growth phenotype as well (Figure 5E).

We conclude that SNC1 is a natural modifier of mkp1 and thus plays a major role in its accession-dependent growth phenotype. Altered SNC1 gene expression, however, is unlikely the cause of the detrimental effects detected in mkp1(Col), but rather the SNC1 signaling pathway is likely sensitized in the absence of MKP1.

**DISCUSSION**

The fitness of all organisms depends largely on their resistance to adverse environmental stress conditions. In particular, sessile plants are inevitably exposed to rapidly changing environmental cues, which have to be recognized and integrated to launch proper responses. Signal transmission and integration often uses MAPK pathways that are regulated by a balance of kinase and phosphatase action. This balance determines the magnitude and duration of MAPK activation and thereby the final biological response. This study demonstrates that (1) MKP1 and PTP1 have redundant functions in regulating plant growth homeostasis by

**Figure 5.** SNC1 Is a Natural Modifier of mkp1.
(A) Photographs of 40-d-old soil-grown plants showing that snc1 largely suppresses the mkp1 growth phenotype (note that the Col and mkp1 photographs are identical to those shown in Figure 2A; all photographs were taken in parallel, under identical conditions). Bars = 2 cm.
(B) to (D) Quantitative RT-PCR analysis of PR1 and PR5 (B), BON1 (C), and SNC1 (D) gene expression. Representative data of three independent experiments are shown. Error bars represent the SD of the technical triplicates.
(E) Photographs of 34-d-old soil-grown plants showing that snc1 partially suppresses the mkp1 ptp1 growth phenotype. Bars = 1 cm.
[See online article for color version of this figure.]
MKP1 and PTP1 Repress Defense Responses

MKP1 plays an important role in the integration and fine-tuning of plant responses to various environmental challenges (Ulm et al., 2002). Under nonstressed conditions, however, growth of the original mkp1 mutant in the Ws background is indistinguishable from that of the wild type; however, the mkp1 mutant has a slightly reduced fertility (Ulm et al., 2001). By contrast, here, we show that loss of MKP1 in the Col background results in a constitutive stress response phenotype causing aberrant growth and development. It should be noted here that the cause for the semidwarf phenotype described for the rice mkp1 mutant might be related (Katou et al., 2007). The constitutive stress response phenotype of mkp1(Col) is exaggerated in the mkp1 ptp1 double mutant, demonstrating an overlapping role of MKP1 and PTP1 as crucial negative regulators of defense responses. However, the presence of MKP1 in the cytoplasm and of PTP1 in the cytoplasm and nucleus indicate that they act in both distinct and overlapping subcellular localizations. The cytosolic localization might indicate an involvement in regulating basal MPK activity levels and/or in early events directly after MPK activation, before translocation of the activated MPKs to the nucleus. Future experiments in which the mutants are complemented with constructs that add artificial nuclear localization and nuclear export signals should be able to address the importance of MKP1 and PTP1 subcellular localization. Notwithstanding this possibility, the functional redundancy between MKP1 and PTP1 seems rather specific, as double mutants of ptp1 or mkp1 with mutants of the other four DSP-type MKPs did not lead to any aberrant phenotype under standard growth conditions (S. Bartels and R. Ulm, unpublished results). Interestingly, an analysis using the Arabidopsis coexpression tool (Manfield et al., 2006) suggests that PTP1 is also the most closely coexpressed gene of MKP1 across a large microarray dataset, in comparison with the other four DSP-type MKPs (see Supplemental Figure 7 online). However, higher-order combinatorial mutants will be required to fully understand the potential functional overlap and specificities among the MAPK inactivating phosphatases. Due to the common MAPK substrates, it will also be of particular interest to determine the potential redundancy between MKP1 and AP2C1 function.

MKP1 and PTP1 have partially overlapping functions as repressors of SA biosynthesis and related defense responses. Consequently, mkp1(Col) and mkp1 ptp1 mutants show enhanced levels of SA, camalexin, PR gene expression, dwarf growth, and early senescence. Interestingly, these phenotypes are largely suppressed by mkp3 and mkp6 mutations. This indicates that MKP3 and 6 are positive regulators of SA-dependent defense responses under negative regulation by MKP1 and PTP1. In agreement, pathogen-induced synthesis of the phytoalexin camalexin was found to be dependent on the activation of MKP3 and MPK6 (Ren et al., 2008). Moreover, pathogen resistance was found to be compromised in MPK6-silenced plants (Menke et al., 2004). Interestingly, the Arabidopsis pathogen P. syringae counteracts the defense-related MPK3 and MPK6 activation by secreting an effector protein with a unique phosphothreonine lyase activity that removes the phosphate group from phosphothreonine to suppress immunity (Zhang et al., 2007a). Altogether, this indicates that MPK3/MPK6 activation is an important node in SA-mediated defense responses, kept under check by MKP1 and PTP1.

Elevated SA-mediated defense responses are usually associated with elevated resistance to biotrophic pathogens. Indeed, we found that mkp1(Col) is more resistant to the bacterial pathogen P. syringae and that this phenotype is suppressed by mkp6, correlating with SA, camalexin, and PR gene expression levels. Interestingly, however, even though these constitutive resistance markers are similarly suppressed in mkp1 mpk3, this double mutant still showed elevated tolerance to P. syringae. This indicates that the elevated resistance of mkp1 is not strictly correlated with its constitutive stress response phenotype but rather with the presence of MKP6. It is also of note that reciprocal inhibition between MPK6 and MPK3 seems to occur (Wang et al., 2008), which could imply that loss of one enhances the activity of the remaining MAPK. An added complexity is apparent by the fact that MKP3, MPK4, and MPK6 protein levels are modulated by SA levels, as indicated by the reduced levels in NahG plants and elevated levels in mkp1(Col) and mkp1 ptp1 (Figure 2B). This observation is in agreement with a recent report on the effect of a functional SA analog (Beckers et al., 2009). Nevertheless, MPK6 is at least partially responsible for the enhanced resistance of mkp1 and likely of mkp1 mpk3, at least in the latter case independent of the constitutively elevated levels of SA and camalexin. However, the exact role of MPK6 in conferring pathogen resistance remains to be determined.

Arabidopsis MKP1 was recently found to be a Ca2+/CaM binding protein, and its phosphatase activity seems to be regulated by this interaction (Lee et al., 2008). Interestingly, Ca2+ signaling was recently found to regulate SA-mediated immunity through the Arabidopsis CaM binding transcription factor SIGNAL RESPONSIVE1 (SR1) that is a repressor of EDS1 expression (Du et al., 2009). Strikingly, the atsr1 null mutant plants exhibit growth and molecular phenotypes associated with elevated SA levels that are very similar to those of mkp1(Col) described here (Du et al., 2009). Thus, it is intriguing to speculate that there is a connection between Ca2+ signaling and MAPK regulation in defense responses that involves CaM-mediated MKP1 regulation, in parallel with SR1 regulation.

The five DSP-type MKPs are potential negative regulators of MAPK pathways, which consist of the 20 MAPks, 10 MAPKKs, and >60 MAPKKKs present in the Arabidopsis genome (Ichimura et al., 2002). Six MAPKKs were linked to the activation of MPK3 and MPK6, namely, MKks 2, 3, 4, 5, 7, and 9 (Asai et al., 2002; Teige et al., 2004; Takahashi et al., 2007; Yoo et al., 2008; Popescu et al., 2009; Zhou et al., 2009). This already complex network was recently extended by an extensive in vitro protein interaction screen to identify MKK6 for activation of MPK3 and 6, and MKK1 for MPK6 (Popescu et al., 2009). Consistent with this multitude of activators and pathways, the stress-activated MPKs
MKK7 overexpression in Arabidopsis phenotypes very much resemble the phenotype of dwarfism and early senescence (Zhou et al., 2009). These MKK9-MPK6 cascade in regulation of leaf senescence, with alterations in combination with SA accumulation, constitutive PR gene transcription, and elevated pathogen resistance (Zhang et al., 2007b). Similarly, a recent report demonstrated a role of the MKK9-MPK3/MPK6 signaling pathways that regulate stress responses and senescence. It is of note that knockouts in MEKK1, MKK1/MKK2, and MPK4 also result in dwarf growth due to a constitutive stress response phenotype and elevated levels of SA (Petersen et al., 2000; Ichimura et al., 2006; Nakagami et al., 2006; Qiu et al., 2008). Similar to mkp1(Col) and mpk1 ptp1 described here, and the MKK9 overexpression phenotypes were also suppressed by the mpk6 mutation (Zhou et al., 2009). Thus, MKP1 and PTP1 likely serve as crucial regulators of the MKK7/MKK9-MPK3/MPK6 pathways in defense responses that regulate stress responses and senescence.

In general, resistance genes are known to rapidly duplicate and mutate to create new specific resistance to constantly evolving pathogens (Noel et al., 1999). The advantage of multiple resistance genes is counterbalanced by potentially detrimental misexpression or misactivation that might lead to autoimmune-type responses. Thus, these rapidly evolving clusters are potent activators of plant defense and need to be tightly regulated. Recently, clusters of TIR-NB-LRR disease resistance genes have been shown to elicit hybrid necrosis, a type of deleterious autoimmune response resulting from incompatibility of Arabidopsis ecotypes. It was thus suggested that such a mechanism might create gene flow barriers in evolution (Bombíles et al., 2007; Alcazar et al., 2009). Our data suggest that MKP1/PTP1-regulated MAPK pathways are likely involved in the regulation of such intra- and interspecific compatibilities, which ensures an optimal balance between growth and pathogen resistance by modulating SA biosynthesis.

METHODS

Plant Material and Growth Conditions

bon1-1, eds1-22 (SALK_071051), mpk3-1 (SALK_151594), mpk4 (SALK_056245), mpk6-2 (SALK_073907), mpk6-3 (SALK_127507), pad4-1 (SALK_073806), mpk6-1 (SALK_118658), snc1-11, and the NahG transgenic lines are all in the Col ecotype (Gaffney et al., 1993; Zhou et al., 1998; Alonso et al., 2003; Yang and Hua, 2004). The mpk1-1 mutation (Ulmi et al., 2001) was introgressed from its Ws background into Col by crossing for at least nine times.

To generate double mutants, single mutants were crossed and the double mutants identified by PCR genotyping in the F2 generation. Plant genomic DNA for PCR analysis was prepared according to Edwards et al. (1991). T-DNA- and gene-specific primers were used as described in Supplemental Table 1 online.

Arabidopsis thaliana seeds were surface-sterilized with sodium hypochlorite and plated on half-strength Murashige and Skoog medium (Duchefa) supplemented with 1% sucrose and 0.5% Phytagel (Sigma-Aldrich). Seeds were stratified for at least 2 d at 4°C and germinated aseptically at 24°C in a standard growth chamber (MLR-350; Sanyo) with a 12-h/12-h light/dark cycle. Standard growth on soil in a phytocam chamber was under a 16-h/8-h light/dark cycle with 21°C/19°C, if not otherwise indicated.

Generation of Transgenic Arabidopsis Lines

The MKP1 and PTP1 coding regions were cloned by the Gateway BP reaction into pDONR207 using primers attB1-MKP1/attB2-MKP1-STOP and attB1-PTP1/attB2-PTP1-STOP (see Supplemental Table 2 online), respectively, and verified by sequencing. Gateway-based cloning was then used to insert PTP1 into the binary destination vector pB7YWG2 and MKP1 into pH7YWG2 (Karimi et al., 2002). Arabidopsis plants were transformed by Agrobacterium tumefaciens using the floral dip method (Clough and Bent, 1998). The resulting transgenic lines described in this work were genetically determined to have the transgene integrated at a single locus.
BiFC Assay and Epifluorescence Microscopy

*Sinapis alba*–based BiFC assays were performed as described previously (Stolpe et al., 2005). A Pro35S:CFP control plasmid was always coombed to identify transformed cells prior to the analysis of YFP fluorescence. PCR fragments for MKP1, PTPI, and MKP coding sequences (including stop codons) were generated with the primers listed in Supplemental Table 2 online and cloned with the Gateway BP reaction into pDONR207 generating entry clones. Gateway LR reaction was then used to clone MKP1 and PTPI into the pE-SPYNE-GW destination vector and the MPKs into pE-SPYCE-GW. The empty vectors used as negative controls were generated by recombination with an empty pENTRY3C clone, which was produced by EcoRI digestion and self-ligation to remove the ccppG gene. The Gateway-compatible BiFC binary vectors pE-SPYNE-GW and pE-SPYCE-GW were kindly provided by Caroline Eco (University of Göttingen).

Transient Expression in *Arabidopsis* Protoplasts and Immunocomplex Kinase Assays

The open reading frames of MPK6 and MKP1 were cloned into the plant expression vector pGREEN and constitutively active MKK4ΔE (with both putative phosphorylation sites changed to Glu residues: T224E and T228E) or to a c-myc epitope (MKP1), as described before (Doczi et al., 2007). Arabidopsis protoplast transient expression assays and protein extraction were performed as described (Ouaked et al., 2003). For immunocomplex kinase assays MPK6 was isolated from the total protein extract using an anti-MPK6–specific antibody incubated for 2 h at 4°C together with Sepharose-A beads. Kinase reactions of the immunoprecipitated MPK6 proteins were performed at 30°C for 30 min in 15 μL of kinase buffer (20 mM HEPES, pH 7.4, 10 mM MgCl2, 5 mM EGTA, and 1 mM DTT) containing 1 μg of MBP, 0.1 mM ATP, and 2 μCi of [γ-32P]ATP. The kinase reactions were stopped by adding 4× SDS loading buffer, and proteins were separated by 15% SDS-PAGE. MBP phosphorylation was analyzed using a phosphor imager and quantified using ImageQuant analysis software (GE Healthcare).

Generation of Antibodies, Immunoprecipitation Assays, and Protein Gel Blot Analysis

Rabbit polyclonal antibodies were generated against synthetic peptides derived from the MPK3 protein sequence (MTTGGGGQYTDFPAEVC, MPK4 (CGFSSGGDQGGSGKGA), and MPK6 (MDGSSQPAADTMTG) and were affinity purified against the peptide (Eurogentec).

For protein gel blot analysis, total cellular proteins (10 μg) were separated by electrophoresis in 10% SDS-polyacrylamide gel and electrophoretically transferred to a polyvinylidene fluoride membrane according to the manufacturer’s instructions (Bio-Rad). We used polyclonal primary antibodies against MPK3, MPK4, MPK6, actin (Sigma-Aldrich), and phospho-p44/42 MAP kinase (Cell Signaling Technologies) and monoclonal anti-GFP (BAbCO), with horseradish peroxidase–conjugated anti-rabbit, anti-goat, and anti-mouse immunoglobulins (DAKO) as secondary antibodies, as required. Signal detection was performed using the ECL Plus Western Detection Kit (GE Healthcare).

Quantitative Real-Time PCR

*Arabidopsis* total RNA was treated with DNasel according to the manufacturer’s specifications (Qiagen). Per PCR reaction, cDNA was synthesized from 25 ng of RNA with a 1:1 mixture of random hexamers and oligo(dT) using the TaqMan reverse transcription reagents kit (Applied Biosystems). Quantitative RT-PCR was performed in a 96-well format using a 7300 real-time PCR system (Applied Biosystems). PCR reactions were performed using the ABsolute SYBR Green Rox Mix Kit following the manufacturer’s instructions (Thermo Scientific). The gene-specific probes and primers were as follows: SNC1 with SNC1_for (5′-AGGAT- TAGATCTGTGTGATGC-3′) and SNC1_rev (5′-CCCTCCTACATTTAGGAAAGC-3′); BON1 with BON1_for (5′-CTCTGCTCTAAGATTCCAA-CAG-3′) and BON1_rev (5′-GAATTGAGTCATCCATGCTCTAC-3′); PR1 with PR1_for (5′-AGAGTGTAGTGGTACGGTGG-3′) and PR1_rev (5′-CTCTGTGAAGCTGTCTTTC-3′); MKP1 with MKP1_for (5′-CAGATCTGCTGATGCATTGG-3′) and MKP1_rev (5′-TGCTAGGAACCTCTGCTTC-3′); and IC51 with IC51_for (5′-CTGGTCTGAAAGCCATAC-3′) and IC51_rev (5′-ATTTGCGCTTGAAGCCATAC-3′). For PR5, TaqMan quantitative RT-PCR was performed using the gene-specific probe 6-FAM-ACAGACTCCTACTTAAAGCATTGCCCT-TAMRA with primers PR5_for (5′-CTTCTTCTCTGTGTCCATAC-3′) and PR5_rev (5′-TCTTTCGACGGAGAGTCT-3′) as described before (Favory et al., 2009), cDNA concentrations were normalized to the 18S rRNA transcript levels as standard using the Eukaryotic 18S rRNA Kit (Applied Biosystems). Average results of three biological replicates, each measured as triplicates, are given.

**SA and Camalexin Extraction and Quantification by HPLC**

Samples (500 mg) were taken from soil-grown 22-d-old plants, combining rosette from three to four plants per sample. SA and camalexin were extracted and quantified by HPLC as previously described (Garcion et al., 2008). Average results from at least three different samples per genotype are given.

**Pseudomonas syringae Growth Assays and Luciferase Imaging**

Plants were maintained at 22°C day/20°C night with 9-h daylength. Inoculation of plants with *P. syringae* pv tomato (Pst) was performed essentially as described (Zipfel et al., 2004). Prior to inoculation, the bioluminescent strain of *Pst* DC3000 containing the LuxCDABE operon (Fan et al., 2008) was streaked onto a King’s B agar containing 50 μg/mL kanamycin and 60 μg/mL rifampicin. After 3 d, bacteria were scraped from the plate, resuspended to a final OD600 = 0.1 in sterile water containing 0.04% Silwet, and sprayed onto the upper surface of leaves. Plants were covered for 24 h postinfection to maintain higher humidity. Luciferase signal from whole rosettes or detached leaves was detected using the Photek HRPCS4 photon detection camera (Photek). Counting of leaf bacteria by serial dilution plating was performed as previously described (Katagiri et al., 2002). Prior to imaging detached leaves or sampling for serial dilution plating was performed as previously described (Favory et al., 2009). cDNA concentrations were normalized to the 18S rRNA Kit (Applied Biosystems). Average results from at least three different samples per genotype are given.

**Accession Numbers**

Sequence data from this article can be found in the EMBL/GenBank data libraries under accession numbers AT1G10210 (MKP1), AT1G59680 (MKP2), AT1G17860 (PTP1), AT1G74170 (ICS1/SID2), AT1G75040 (PR5), AT2G14610 (PR1), AT2G43790 (MPK6), AT3G45640 (MKP3), AT3G55270 (MKP1), AT4G01370 (MPK4), AT4G16890 (SNC1), and AT5G61900 (BON1).

**Supplemental Data**

The following materials are available in the online version of this article.

**Supplemental Figure 1.** Characterization of the *ptp1-1* T-DNA Insertion.

**Supplemental Figure 2.** Comparison of *mkp1* (Ws) versus *mkp1* (Col), Complementation of *mkp1* (Col) with a Pyo-MKP1 Transgene, and *mkp1-1* Coexpression (c5s) Line.
Supplemental Figure 3. Growth Phenotypes of mkp1(Col) and mkp1 ptp1 Appear Rather Late in Development.

Supplemental Figure 4. Suppression of the mkp1(Col) Growth Phenotype by the mkp6-3 Allele.

Supplemental Figure 5. Levels of Free and Conjugated Salicylic Acid.

Supplemental Figure 6. Suppression of mkp1(Col) and mkp1 ptp1 Phenotypes by Elevated Temperatures (28°C).

Supplemental Figure 7. Correlation Scatterplot of MKP1 and PTP1 Coexpression Compared with IBR5, PHS1, MKP2, and DsPTP1.

Supplemental Figure 8. mkp6 Mutation Does Not Suppress the mkp4 Dwarf Growth Phenotype.

Supplemental Table 1. Primer Sequences Used for Mutant Genotyping.

Supplemental Table 2. Primer Sequences Used for Molecular Cloning.

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